

STUDIES ON AMYLASE SYNTHESIS BY THERMOPHILIC *Bacillus* sp. ISOLATED FROM REFUSE DUMP AND ITS ACTION ON STARCHY WASTE MATERIALS.

BY

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2014



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Action on Starchy Waste Materials

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This research project was supervised by me and approved in accordance with the partial fulfilment for the award of Master of Science (M. Sc.) degree in Microbiology, Obafemi Awolowo University, Ile –Ife, Nigeria.

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DEDICATION

With humility and profound gratitude I dedicate this project to my supervisor; Revd. (Dr.) M. K. Bakare for both intellectual grooming and financial support during the course of this work.

I also dedicate this research to my mentor Dr. (Mrs.) A. O. Oluduro for her persistent provisions and guidance before, during and after this work. God will bless your glorious home and career. I gladly dedicate this thesis to Miss Yetunde Helen Abioye for your love, support and unfathomable hospitality throughout the course of my M. Sc. most especially during the practicals.

To my ineffable parents Mr. and Mrs. Samson Ayodele Omoboye and all my family members for prayers and support at all times, I heartily dedicate this thesis.



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TABLE OF CONTENTS

Title		P	age
Title pa	ge		i
Authori	zation to Copy		ii
Certific	ation		iii
Dedicat	ion		iv
Acknow	ledgements		v
Table o	f Contents		vii
List of T	ables		xiv
List of F	List of Figures xv		
List of P	lates		xvii
Abstract			xviii
СНАРТЕ	RONE	1	
1.0	Introduction		1
1.1	Starch	1	
1.2	Enzymes and Alpha- Amyl ase	2	
1.3	Industrial Application of Amylases		3
1.4	Justification		4



1.5	The Specific Objectives	5	
CHAPTER	RTWO	6	
2.0	Literature Review		6
2.1	The Genus Bacillus		6
2.1.1	Industrial Significance of <i>Bacillus</i> spp.	6	
2.1.2	Use of Bacillus spp. as Model Organism		6
2.2	Microbial Alpha-amylase		7
2.2.1	Bacterial Amylases		8
2.2.2	Fungal Amylases		10
2.2.3	Purification of Alpha-amylase	11	
2.2.4	Industrial Application of Microbial Alpha-amylase and Other Amylases	12	
2.2.4.1	Starch Conversion		12
2.2.4.2	Detergent Industry		13
2.2.4.3	Fuel Alcohol Production		14
2.2.4.4	Food Industry	14	
2.2.4.5	Textile Industry		15
2.2.4.6	Paper Industry		16
2.2.4.7	Liquefaction	16	

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2.3	Waste Management		16
2.3.1	Solid Waste Management		17
2.3.2	Food and Yard Wastes		19
2.3.3	Organic Wastes		19
CHAPTER	THREE	2	20
3.0	Materials and Methods		20
3.1	Materials		20
3.2	Methods		20
3.2.1	Study Location		21
3.2.2	Collection of Samples and Culturing	21	
3.2.3	Identification of Isolates		21
3.2.3.1	Cultural Characteristics		22
3.2.3.2	Gram Staining		22
3.2.3.3	Spore Staining		22
3.2.3.4	Biochemical Identification of the Isolates	23	
3.2.3.4.1	Catalase Test	23	
3.2.3.4.2	Citrate Test	23	
3.2.3.4.3	Indole Test	24	
3.2.3.4.4	Methyl Red and Voges Proskauer (MR-VP) Test	24	



3.2.3.4.5	Sugar Fermentation	24	
3.2.3.4.6	Motility Test	25	
3.2.3.4.7	Oxidase Test	25	
3.2.3.4.8	Urease Test	25	
3.2.3.4.9	Starch Hydrolysis	2	26
3.2.3.4.10	Hydrogen Sulphide Production		26
3.2.3.4.11	Growth at 6.5 % Sodium Chloride (NaCl) Concentration		26
3.2.3.5	Molecular Characterization of the Bacteria DNA	27	
3.2.3.5.1	Extraction of DNA Using Cetyltrimethylammonium Bromide (CTAB)		
	Method		27
3.2.3.5.2	DNA Electrophoresis	27	
3.2.3.5.3	Polymerase Chain Reaction (PCR) Analysis Using 16S Primer	28	
3.2.3.5.4	Purification of PCR Products	28	
3.2.3.5.5	PCR product for Sequencing	28	
3.2.3.5.6	Purification of PCR Sequencing Products	29	
3.2.3.5.7	Preparation of Sample for Gene Sequencer (ABI machine)	29	
3.2.3.5.8	Blasting of the Gene Sequence		29
3.2.3.6.	Methods for Extraction of the Raw Starches	30	
3.2.3.6.1	Extraction of Starch from Sorghum	30	



3.2.3.6.2	Extraction of Starch from White Yam	30	
3.2.3.6.3	Extraction of Starch from Cassava	31	
3.2.3.6.4	Extraction of Starch from Corn		31
3.2.3.6.5	Preparation of Cassava Flour	31	
3.2.3.7	Bacillus sp. RD24 Alpha Amylase Enzyme	32	
3.2.3.7.1	Alpha Amylase Production and Extraction	32	
3.2.3.7.1.1	Alpha Amylase Assay		32
3.2.3.7.1.2	Protein Assay		33
3.2.3.7.2	Growth and Enzyme Production		33
3.2.3.7.3	Optimization of Alpha Amylase Production	33	
3.2.3.7.3.1	Determination of Optimum pH for Alpha-amylase production		33
3.2.3.7.3.2	Determination of Optimum Temperature for Alpha Amylase Production	33	
3.2.3.7.3.3	Effect of Soluble Starch Concentration on Enzyme Production		34
3.2.3.7.3.4	Effect of Some Nitrogen Sources on Enzyme Production	34	
3.2.3.3.7.5	Effect of Some Carbon sources on Enzyme Production		34
3.2.3.7.4	Effect of Some Raw Starch Sources on Enzyme Production	35	
3.2.3.7.5	Enzyme Purification	34	
3.2.3.7.5.1	Ammonium Sulphate Precipitation	35	
3.2.3.7.5.2	Dialysis		36



3.2.3.7.5	3 Enzyme Purification of the Cell-free Supernatant by Ion – Exchange		
	Chromatography on CM Sepharose CL-6B		36
3.2.3.8	Determination of Native Molecular Weight	36	
3.2.3.9	Effects of pH on Activity of Partially Purified Bacillus sp.		
	RD24 α - amylase after 20 minutes of incubation	37	
3.2.3.10	Effects of Temperature on Activity of Partially Purified Bacillus sp.		
	RD24 α-amylase		37
3.2.3.11	Effects of Metal Ion on Partially Purified Bacillus sp. RD24 Alpha		
	Amylase		37
3.2.3.12	Effects of EDTA on the Partially Purified Bacillus sp. RD24 Alpha		
	Amylase		38
3.2.3.13	Effect of Partially Purified Bacillus sp. RD24 Alpha Amylase on		
	Raw Starches		38
3.2.3.15	Determination of Kinetic Parameters		38
CHAPTER	FOUR	40	
4.0 R	ESULTS		40
4.1 ls	solation of Amylolytic Bacteria	40	
4.2 lo	dentification of Amylolytic Bacteria		40
4.3 G	rowth and Alpha Amylase Production	45	



4.4	Optimization of Production Conditions for Alpha Amylase Production	45	
4.4.1	Optimization of pH and Temperature for Alpha Amylase Production by		
	Bacillus licheniformis RD24		45
4.4.2	The Percentage Starch Composition on Alpha Amylase Production by		
	Bacillus licheniformis RD24	\mathcal{A}	48
4.4.3	Effect of Nitrogen Sources on the Production of Alpha Amylase by Bacillus		
	licheniformis RD24		48
4.4.4	Effect of Carbon Sources on the Production of Alpha Amylase by Bacillu	s	
	licheniformis RD24		48
4.5	Effect of Raw Starches on the Production of Alpha Amylase by		
	Bacillus licheniformis RD24		51
4.6	Purification of Alpha - amylase Using Ion Exchange Column Chromatography		
	on CM Sepharose CL-6B	48	
4.7	Bacillus licheniformis RD24 Native Molecular Weight	52	
4.8	Effect of pH on the Activity of Partially Purified Bacillus licheniform is RD24		
	α-amylase after 20 minutes of Incubation		52
4.9	Effect of pH on the Activity of Partially Purified Bacillus licheniformis		
	RD24 Alpha Amylase at Different pH after 60 minutes of Incubation	52	
4.10	Effect of Temperature on the Activity of Partially purified Bacillus licheniformis		



	RD24 α-amylase.		56
4.11	Effect of Metal Ions on <i>Bacillus licheniformis</i> RD24 Alpha Amylase Activity	56	
4.12	Effect of EDTA on Purified Bacillus licheniformis RD24 Alpha Amylase		
	Activity	59	
4.13	Effect of Purified Bacillus licheniformis RD24 Alpha Amylase on Raw Starches	59	
4.14	Bacillus licheniformis RD24 Kinetic Parameters	63	
CHAPTI	ER FIVE	66	
5.0	DISCUSSION, CONCLUSION AND RECOMMENDATION 66		
5.1	Discussion		66
5.2	Conclusion		74
5.3	Recommendation		75
REFERE	INCES	76	
APPENI	DICES	94	
APPENI	DIX I: Preparation of Culture Media		94
APPENI	DIX II : Preparation of Reagents		104
APPENI	DIX III: Standard Curves	106	
APPENI	DIX IV: Results from Figures	109	



TABLES OF CONTENTS

Table	Title		
Page			
4.1	Morphological Characteristics of the Isolates		40
4.2	Gram's Staining, Spore Staining and Biochemical Characteristics of the	2	
ls	solates	41	
4.3	Frequency and Percentage Distribution of the Amylolytic Bacteria		42
4.4	Sequences Producing Significant Alignments with Bacillus licheniformis		
I	RD24		44
4.5	Summary of Purification Protocols of Alpha-amylase Obtained		
08	from Bacillus licheniformis RD24	54	



LIST OF FIGURES

 4.1 Growth and Enzyme Production Curve at 45 °C and pH for 48 h 4.2 Effect of pH on the Production of Bacillus licheniformis RD24 Crude α-amylase 4.3 Effect of Temperature on the Production of Bacillus licheniformis RD24 Crude α- amylase 4.4 Effect of Percentage (%) Starch Composition on the Production of Bacillus licheniformis RD24 Crude α-amylase 4.9 4.5 Effect of Nitrogen Source on the Production of Bacillus licheniformis RD24 Crude α-amylase 4.9 4.6 Effect of Carbon Source on the Production of Bacillus licheniformis RD24 Crude α-amylase 4.9 4.7 Effect of Raw Starches on the Production of Bacillus licheniformis RD24 Crude α-amylase 4.7 Effect of Raw Starches on the Production of Bacillus licheniformis 4.8 Elution profile of Bacillus licheniformis RD24 α-amylase on CM 	Figu	ure Title	Page
α-amylase 47 4.3 Effect of Temperature on the Production of Bacillus licheniformis RD24 Crude α- amylase 47 4.4 Effect of Percentage (%) Starch Composition on the Production of Bacillus licheniformis RD24 Crude α-amylase 49 4.5 Effect of Nitrogen Source on the Production of Bacillus licheniformis 49 4.6 Effect of Carbon Source on the Production of Bacillus licheniformis 49 4.6 Effect of Carbon Source on the Production of Bacillus licheniformis 50 4.7 Effect of Raw Starches on the Production of Bacillus licheniformis 50 4.7 Effect of Raw Starches on the Production of Bacillus licheniformis 50 4.7 Effect of Raw Starches on the Production of Bacillus licheniformis 50 4.7 Effect of Raw Starches on the Production of Bacillus licheniformis 50 4.7 Effect of Raw Starches on the Production of Bacillus licheniformis 51	4.1	Growth and Enzyme Production Curve at 45 °C and pH for 48 h	46
 4.3 Effect of Temperature on the Production of Bacillus licheniformis RD24 Crude α - amylase 47 4.4 Effect of Percentage (%) Starch Composition on the Production of Bacillus licheniformis RD24 Crude α - amylase 49 4.5 Effect of Nitrogen Source on the Production of Bacillus licheniformis RD24 Crude α - amylase 49 4.6 Effect of Carbon Source on the Production of Bacillus licheniformis RD24 Crude α - amylase 49 4.6 Effect of Carbon Source on the Production of Bacillus licheniformis AD24 Crude α - amylase 49 4.6 Effect of Carbon Source on the Production of Bacillus licheniformis AD24 Crude α - amylase 49 50 51 	4.2	Effect of pH on the Production of Bacillus licheniformis RD24 Crude	
RD24 Crude α - amylase 47 4.4 Effect of Percentage (%) Starch Composition on the Production of Bacillus licheniformis RD24 Crude α - amylase 49 4.5 Effect of Nitrogen Source on the Production of Bacillus licheniformis RD24 Crude α - amylase 49 4.6 Effect of Carbon Source on the Production of Bacillus licheniformis RD24 Crude α - amylase 49 4.6 Effect of Carbon Source on the Production of Bacillus licheniformis RD24 Crude α - amylase 50 4.7 Effect of Raw Starches on the Production of Bacillus licheniformis RD24 Crude α - amylase 50 A.7 Effect of Raw Starches on the Production of Bacillus licheniformis RD24 Crude α - amylase 50		α-amylase	47
4.4Effect of Percentage (%) Starch Composition on the Production of Bacillus licheniformis RD24 Crude α-amylase494.5Effect of Nitrogen Source on the Production of Bacillus licheniformis RD24 Crude α-amylase494.6Effect of Carbon Source on the Production of Bacillus licheniformis RD24 Crude α-amylase504.7Effect of Raw Starches on the Production of Bacillus licheniformis RD24 Crude α-amylase50	4.3	Effect of Temperature on the Production of Bacillus licheniformis	2/
Bacillus licheniformis RD24 Crude α-amylase494.5Effect of Nitrogen Source on the Production of Bacillus licheniformisRD24 Crude α-amylase494.6Effect of Carbon Source on the Production of Bacillus licheniformisRD24 Crude α-amylase504.7Effect of Raw Starches on the Production of Bacillus licheniformisRD24 Crude α-amylase504.7Effect of Raw Starches on the Production of Bacillus licheniformisA.7Effect of Raw Starches on the Production of Bacillus licheniformisA.7Effect of Raw Starches on the Production of Bacillus licheniformisA.7Effect of Raw Starches on the Production of Bacillus licheniformisA.7Effect of Raw Starches on the Production of Bacillus licheniformisA.7Effect of Raw Starches on the Production of Bacillus licheniformisA.7Effect of Raw Starches on the Production of Bacillus licheniformisA.7Effect of Raw Starches on the Production of Bacillus licheniformisA.7Effect of Raw Starches on the Production of Bacillus licheniformisA.7Effect of Raw Starches on the Production of Bacillus licheniformisA.7Effect of Raw Starches on the Production of Bacillus licheniformisA.7Effect of Raw Starches on the Production of Bacillus licheniformisA.7Effect of Raw Starches on the Production of Bacillus licheniformisA.7Effect of Raw Starches on the Production of Bacillus licheniformisA.7Effect of Raw Starches on the Production of Bacillus licheniformisA.7Effect of Raw Starches on the Production of Bacillus licheniformis		RD24 Crude α- amylase	47
 4.5 Effect of Nitrogen Source on the Production of Bacillus licheniformis RD24 Crude α-amylase 49 4.6 Effect of Carbon Source on the Production of Bacillus licheniformis RD24 Crude α-amylase 4.7 Effect of Raw Starches on the Production of Bacillus licheniformis A.7 Effect of Raw Starches on the Production of Bacillus licheniformis A.7 Effect of Raw Starches on the Production of Bacillus licheniformis A.8 Effect of Raw Starches on the Production of Bacillus licheniformis A.9 Effect of Raw Starches on the Production of Bacillus licheniformis 	4.4	Effect of Percentage (%) Starch Composition on the Production of	
RD24 Crude α-amylase494.6Effect of Carbon Source on the Production of Bacillus licheniformisRD24 Crude α-amylase504.7Effect of Raw Starches on the Production of Bacillus licheniformisRD24 Crude α-amylase51		<i>Bacillus licheniformis</i> RD24 Crude α-amylase	49
4.6Effect of Carbon Source on the Production of Bacillus licheniformisRD24 Crude α-amylase504.7Effect of Raw Starches on the Production of Bacillus licheniformisRD24 Crude α-amylase51	4.5	Effect of Nitrogen Source on the Production of Bacillus licheniformis	
RD24 Crude α-amylase 50 4.7 Effect of Raw Starches on the Production of Bacillus licheniformis RD24 Crude α-amylase 51		RD24 Crude α-amylase	49
4.7Effect of Raw Starches on the Production of Bacillus licheniformisRD24 Crude α-amylase51	4.6	Effect of Carbon Source on the Production of Bacillus licheniformis	
RD24 Crude α-amylase 51		RD24 Crude α-amylase	50
J.	4.7	Effect of Raw Starches on the Production of Bacillus licheniformis	
4.8 Elution profile of <i>Bacillus licheniformis</i> RD24 α-amylase on CM		RD24 Crude α- amylase	51
	4.8	Elution profile of Bacillus licheniformis RD24 α -amylase on CM	
Sepharose CL-6B 53		Sepharose CL-6B	53
4.9 Effect of pH on Partially Purified <i>Bacillus licheniformis</i> RD24	4.9	Effect of pH on Partially Purified Bacillus licheniformis RD24	
Enzyme Activity after 20 minutes of incubation 55		Enzyme Activity after 20 minutes of incubation	55

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4.10	Effect of pH on Partially Purified Bacillus licheniformis RD24 α -amylase	in	
	Buffers of Different pH after 20 minutes of incubation	55	
4.11	Effect of Temperature on Partially Purified Bacillus licheniformis RD24 E	inzyme	
	Activity	57	
4.12	Effect of Na ⁺ on the Activity of Partially Purified <i>Bacillus licheniformis</i>	2	
	RD24 α -amylase at 70 °C and pH 8		58
4.13	Effect of Mg ²⁺ on the Activity of Partially Purified Bacillus licheniformis		
	RD24 α -amylase at 70 °C and pH 8	58	
4.14	Effect of Ca ²⁺ on the Activity of Partially Purified <i>Bacillus licheniformis</i>		
	RD24 α -amylase at 70 °C and pH 8		58
4.15	Effect of Al ³⁺ on the Activity of Partially Purified <i>Bacillus licheniformis</i>		
	RD24 α -amylase at 70 °C and pH 8		58
4.16	Effect of EDTA on the Activity of Partially Purified Bacillus licheniformis		
1	RD24 α -amylase at 70 °C and pH 8	60	
4.17	Effect of Partially Purified Bacillus licheniformis RD24 α -amylase on		
	Gelatinized Raw Starch at 70 °C and pH 8	61	
4.18	Effect of Partially Purified Bacillus licheniformis RD24 α -amylase on		
	Ungelatinized Raw Starch at 70 $^\circ C$ and pH 8	62	
4.19	Effect of Soluble Starch on Partially Purified α -amylase	64	



4.20 Lineweaver-Burk Plot for the Determination of Kinetic Parameters of Partially

Purified α-amylase Produced by *Bacillus licheniformis* RD24 65



LIST OF PLATES

PLATE	TITLE PAGE	
1.1	3D Structure of Human Amylase	8
4.1	Agarose Electrophoresis of Polymerase Chain Reaction (PCR)	43
	product of <i>Bacillus licheniformis</i> RD24	



ABSTRACT

This study isolated and characterized α -amylase-producing thermophilic bacteria from a refuse dumpsite as well as characterized the partially purified α -amylase produced by the isolated bacteria. This was with a view to obtaining a thermostable α -amylase-producing bacteria capable of digesting raw starches.

Four soil samples were collected at different dumpsites in Ile-Ife, Osun State, Nigeria. Serially diluted inocula were plated on nutrient agar in order to isolate bacteria for their α -amylase activity. The isolated bacteria were identified by morphological and biochemical characterization while the bacteria of interest was identified by molecular analysis using 16S rRNA gene sequencing. The bacterium with the highest α -amylase activity was selected for enzyme production, purification and characterization. The optimal conditions for α -amylase secretion by the bacterium were determined by varying the pH, temperature, percentage soluble starch, nitrogen sources and carbon sources. Sources of raw starches were also varied. The enzyme was partially purified by ion exchange chromatography on CM Sepharose CL-6B. The molecular weight of the enzyme was determined using gel filtration on Sephadex G-100. Kinetic parameters (K_m and V_{max}) of the purified enzyme, effects of temperature, pH, metal ions and ethylenediaminetetra acetic acid (EDTA) were studied.

The isolated and identified bacteria were *Bacillus alvei* (40%) *Bacillus licheniformis* (40%) and *Bacillus brevis* (20%) while *Bacillus licheniformis* RD24 was identified by 16S rRNA gene sequencing. The peak of amylase productivity was at 20 h of incubation (925 μ g/ml/min). The optimum pH and temperature for the production of *Bacillus licheniformis* RD24 α -amylase were 7 (with 150 ± 1.33 μ g/ml/min) and 45 °C (with 58 ± 1.66 μ g/ml/min enzyme activity) respectively. One percent (1%) starch composition of the enzyme production medium gave highest enzyme activity of 102 ± 5.3 μ g/ml/min.



Peptone gave an enzyme activity of 165 ± 8.97 µg/ml/min and yeast extract gave 52.26 ± 2.86 µg/ml/min. Starch gave the highest activity of 33 ± 4.98 Units/ml followed by lactose (32 ± 2.99 Units/ml) and melibiose (27 ± 2.99 µg/ml/min). Of the raw starches, cassava flour gave the highest specific activity of 72 ± 0.07 Units/mg protein, while sorghum starch gave the lowest specific activity 5 ± 1.52 Units/mg protein. The specific activity of the partially purified *Bacillus licheniformis* RD24 α -amylase for starch was 1.634 Units/mg protein with a purification fold of 4.76. The partially purified *Bacillus licheniformis* RD24 α -amylase had a molecular weight of 50 kDa. The V_{max} and K_m of the partially purified *Bacillus licheniformis* RD24 α -amylase with soluble starch as substrate were 4654 ± 108 Units/mg protein and 79.11 ± 1.84 mg/ml respectively. The optimum pH and temperature of partially purified *Bacillus licheniformis* RD24 α -amylase were 8.0 and 70 °C respectively. Sodium ion had stimulatory effect on the enzyme while Mg²⁺, Ca²⁺ and Al³⁺ inhibited the enzyme. EDTA inhibited the enzyme at all concentrations.

The study concluded that α -amylase synthesized by *Bacillus licheniformis* RD24 had a unique characteristic of thermostability and ability to withstand alkaline pH.



CHAPTER ONE

INTRODUCTION

1.1 Background to the Study

Starch is the most abundant material reservoir in superior vegetable beings destined to nourishment. It is a basic source of chemical energy for life sustenance, which is continuously renewed by sun. Starch production in the earth was estimated to be in the order of 2.0 x 10^{10} tonnes/year, which corresponds to about 80 % of total food production worldwide (Sarikaya *et al.*, 2000). In the most developed tropical countries, where agricultural products like tubers and cereals are abundant, a valuable fraction of starch is yearly lost because of different causes, among which waste and inadequate storage account for (Okolo *et al.*, 1995). Therefore, the conversion of agricultural raw materials into higher added-value products, by enzymatic saccharification, can represent an effective strategy of resources conservation (Okolo *et al.*, 1995). Starch is an abundant carbon source in nature, and α -amylase (1, 4- α -D-glucanohydrolase; EC 3.2.1.1), which hydrolyzes α -1,4-glucosidic linkage in starch-related molecules, is one of several enzymes involved in starch degradation (Verma *et al.*, 2011).

Starch is the most common carbohydrate in the human diet and is contained in many staple foods. The major sources of starch intake worldwide are the cereals (rice, wheat, and maize) and the root vegetables (sweet potatoes and cassava) (Eliasson, 2004). Many other starchy foods are grown, some only in specific climates, including acorns, bananas, barley, breadfruit, colacasia, katakuri, millet, oats, polynesian arrowroot, sago, sorghum, sweet potatoes,



rye, chestnuts and yams, and many kinds of beans, such as favas, lentils, mung beans, peas, and chickpeas (Eliasson, 2004).

1.2 Enzymes and Alpha Amylase

Enzymes are among the most important products acquired for human needs in the areas of industrial, environmental and food biotechnology through microbial sources. Alpha amylase is a hydrolytic enzyme and in recent years, interest in its microbial production has increased dramatically due to its wide spread use in food, textile, baking and detergent industries (Asgher *et al.*, 2007). Starch-degrading enzymes like amylase have received great deal of attention because of their perceived technological significance and economic benefits (Buzzini and Martini, 2002; Oyeleke and Oduwole, 2009). Amylases are hydrolases that function by the breakdown or hydrolysis of starch into reducing fermentable sugars, mainly maltose and reducing non fermentable or slowly fermentable dextrins (Oyeleke *et al.*, 2010). Amylases are hydrolyzing enzyme in function which causes hydrolysis of starch molecules. In biotechnology amylases are of the most important enzymes used (Aneja, 2003; Burhan *et al.*, 2003).

Alpha amylases (endo - 1, $4 - \alpha - D$ - glucan glucanohydrolase, E.C. 3.2.1.1) are extracellular endo-enzymes that randomly cleave the 1, 4 α -linkage between adjacent glucose units in the linear amylose chain and ultimately generate glucose, maltose and maltotriose units. Among various extracellular enzymes, α -amylase ranks first in terms of commercial exploitation (Babu and Satyanarayana, 1993) and accounts for 12 % of the sales volume of the world enzyme market (Baysal *et al.*, 2003). Spectrum of applications of α -amylase has widened in many sectors such as clinical, medicinal and analytical chemistry. Besides their use in starch saccharification, they also find applications in bakery, brewery, detergent, textile, paper and distilling industry



(Ramachandran *et al.*, 2004). The enzyme is extensively used in many industries including starch liquefaction, brewing, food, paper, textile and pharmaceuticals (Arikan, 2008).

Alpha amylases (1, 4 α -D-glucan glucohydrolase, E.C. 3.2.1.1) are extra cellular enzymes that break down the internal α -1, 4 linkages in starch to form glucose, maltodextrins and maltose. Industrially, α -amylase plays a vital role in starch liquefaction, brewing and food industries (Akpan *et al.*, 2004; Thippeswamy *et al.*, 2006; Gangadharan *et al.*, 2008; Rajagopalan and Krishnan, 2008; Rasiah and Rehm, 2009). *Bacillus* species such as, *B. subtilis, B. amyloliquefaciens, B. stearothermophilus* and *B. licheniformis* are well known potential producers of α -amylase (Gangadharan *et al.*, 2006; Rasiah and Rehm, 2009).

Alpha-amylase has been obtained from several fungi, yeast, bacteria and actinomycetes; however enzymes from fungi and bacteria sources have dominated applications in industrial sectors (<u>Pandey *et al.*</u>, 2000). Evidences of amylase in yeast, moulds and bacteria have been reported and their properties documented (Buzzini and Martini, 2002; Oyeleke and Oduwole, 2009).

1.3 Industrial Application of Amylases

Food and beverage industries employ β -amylase [EC. 3.2.1.2] to convert starch into maltose solutions (Fogarty and Kelly, 1990). Amylases from various fungal and bacterial species have been studied in a great detail and they have been found to be a very good source for amylases production (Khan and Briscoe, 2011).

Bacteria belonging to the genus *Bacillus* have been widely used for the commercial production of thermostable α -amylase (Kubrak *et al.*, 2010). Studies on bacteria amylase especially in the developing countries have concentrated mainly on *Bacillus* spp probably because of the simple nature and nutritional requirements of this organism (Omemu *et al.*, 2005; Ajayi and Fagade, 2006; Oyeleke and Oduwole, 2009). *Bacillus* species are heterogeneous forms



of organisms and they are very versatile in the adaptability to the environment (Aqeel and Umar, 2010).